# Original

Marina Peñuelas¹ Francisco Javier Candel¹ Clara Lejarraga¹ Laura López-González¹ Jose Manuel Viñuela-Prieto¹ Diego López de Mendoza² Activity of linezolid and tedizolid against clinical isolates of methicillin-resistant and methicillin and linezolid resistant *Staphylococcus aureus*: an *in vitro* comparison

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# **ABSTRACT**

Introduction. The prevalence of methicillin-resistant *Staphylococcus aureus* (MRSA) in Spain is approximately 20-30%. However, resistance to linezolid is rare, and the main reports are from nosocomial outbreaks. The objective of the present study was to compare the *in vitro* susceptibility of linezolid with that of tedizolid against MRSA isolates and methicillin- and linezolid-resistant isolates (MLRSA) mediated by the *cfr* gene.

Material and methods. The *in vitro* susceptibility of linezolid and tedizolid was determined using the E-test with 18 MRSA strains and 18 *cfr*-mediated MLRSA strains obtained from clinical isolates in the microbiology service of a tertiary university hospital.

**Results.** All MRSA strains were susceptible to both antibiotics. Analysis of the MRSA isolates revealed that the  $MIC_{50}$  and  $MIC_{90}$  of linezolid were 1.5 and 2 mg/L, respectively; those of tedizolid were 0.25 and 0.4 mg/L. The  $MIC_{50}$  and  $MIC_{90}$  of tedizolid remained at 0.75 and 1 mg/L against the MLRSA strains ( $MIC_{90} \ge 8$  mg/L).

**Conclusions.** Both for MRSA and for MLRSA, the MICs obtained for tedizolid were at least 2 dilutions lower than those of linezolid, thus demonstrating between 2 and 4 times greater activity *in vitro* than linezolid.

Key words: Methicillin-resistant *Staphylococcus aureus*, linezolid-resistant *Staphylococcus aureus*, linezolid, tedizolid, cfr gene.

Actividad comparativa *in vitro* entre linezolid y tedizolid frente a aislados clínicos de *Staphylococcus aureus* resistentes a meticilina y aislados resistentes también a linezolid

#### **RESUMEN**

**Introducción.** La prevalencia de *Staphylococcus aureus* resistente a la meticilina (SARM) se sitúa en España en torno al 20 y el 30%. Sin embargo, la resistencia a linezolid se reporta de forma excepcional, salvo en algunos brotes nosocomiales. El objetivo de nuestro estudio fue realizar un análisis comparativo de la sensibilidad *in vitro* de linezolid y tedizolid frente a aislados de SARM, así como frente a otros también resistentes a linezolid (SARLM) mediados por el gen *cfr.* 

**Material y métodos.** Se determinó la sensibilidad *in vitro* a linezolid y tedizolid mediante la técnica de E-test a 18 cepas SARM y a otras 18 que además presentaban resistencia a linezolid (SARLM) mediadas por el gen *cfr*, procedentes de aislados clínicos en el Servicio de Microbiología de un Hospital terciario Universitario.

**Resultados.** Todas las cepas de SARM fueron sensibles a ambos antibióticos. Analizando los aislados clínicos de SARM, las  $\text{CMI}_{50}\text{-CMI}_{90}$  de linezolid fueron 1,5 y 2 mg/L respectivamente y en el caso de tedizolid de 0,25 y 0,4 mg/L. Frente a las cepas de SARLM ( $\text{CMI}_{90} \geq 8 \text{ mg/L}$ ) las  $\text{CMI}_{50}\text{-CMI}_{90}$  de tedizolid se mantuvieron en 0,75 y 1 mg/L.

**Conclusiones.** Tanto en el caso de las SARM como en el de las SARLM, las CMI obtenidas con tedizolid resultaron ser de al menos dos diluciones más bajas, demostrando entre 2 y 4 veces mayor actividad *in vitro* que linezolid.

Palabras Clave: Staphylococcus aureus resistente a meticilina, Staphylococcus aureus resistente a linezolid, linezolid, tedizolid, cfr gene.

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# **INTRODUCTION**

Resistance to linezolid is increasingly frequent in the hospital environment. This resistance is the result of a genetic change at the binding site of linezolid, namely, 23S ribosomal RNA (rRNA). G2576T is the chromosomal mutation that most commonly leads to this genetic change. Given that all *Staphylococcus* species have between 4 and 7 copies of the gene that codes for 23S rRNA, several mutated copies are necessary before resistance to the drug is expressed, a circumstance that is unlikely to arise unless the patient receives prolonged treatment. Therefore, resistance of *Staphylococcus aureus* to oxazolidinones is less frequent than to other antibiotics, such as methicillin<sup>1</sup>.

Resistance to linezolid mediated by the *cfr* gene, while less common than resistance caused by G2576T, could have a greater clinical impact, since its location in the plasmid potentially facilitates its dissemination in outbreaks. LaMarre et al<sup>2</sup> and Locke et al<sup>3</sup> showed the low *in vitro* fitness cost for *Staphylococcus aureus* of acquiring and expressing this plasmid gene, thus fa-

Table 1

cilitating its propagation independently of antibiotic pressure<sup>2,3</sup>. The *cfr* gene codes for a methyltransferase involved in the methylation of 23S rRNA, thus preventing binding to linezolid as a result of the steric hindrance generated. However, this does not affect the binding capabilities of tedizolid<sup>4,5</sup>.

In the present study, we compare the *in vitro* activity of linezolid with that of tedizolid against strains of methicil-lin-resistant *S. aureus* (MRSA) and *cfr*-mediated methicillin-and linezolid-resistant (MLRSA) obtained during a nosocomial outbreak at our center<sup>6</sup>.

#### MATERIAL AND METHODS

We obtained a total of 18 MRSA and 18 MLRSA strains from clinical samples received in the clinical microbiology department. The MRSA samples were isolated from exudates (9), blood cultures (5), and biopsy samples (4); the MLRSA samples were obtained from respiratory samples (7), blood cultures (5), vascular catheters (2), exudates (3), and cerebrospinal fluid (1). The minimum inhibitory concentration (MIC) was determined

1 2 3 4 5 6 7 8 9 10	1 2 1 1.5 1 0.75 1.5 2	Tedizolid  0.19  0.5  0.125  0.25  0.19  0.19  0.25  0.5	1 2 3 4 5 6 7 8 8	Linezolid  8 6 8 256 128 96 48	7edizolid 0.25 0.5 0.38 0.75 0.75 0.75 0.75
2 3 4 5 6 7 8 9 10	2 1 1.5 1 1 0.75 1.5 2	0.5 0.125 0.25 0.25 0.19 0.19	2 3 4 5 6 7 8	6 8 256 128 96 48	0.5 0.38 0.75 0.75 0.75
3 4 5 6 7 8 9 10	1 1.5 1 1 0.75 1.5	0.125 0.25 0.25 0.19 0.19 0.25	3 4 5 6 7 8	8 256 128 96 48	0.38 0.75 0.75 0.75 0.75
4 5 6 7 8 9 10	1.5 1 1 0.75 1.5 2	0.25 0.25 0.19 0.19 0.25	4 5 6 7 8	256 128 96 48	0.75 0.75 0.75 0.75
5 6 7 8 9 10	1 1 0.75 1.5	0.25 0.19 0.19 0.25	5 6 7 8	128 96 48	0.75 0.75 0.75
6 7 8 9 10 11	1 0.75 1.5 2	0.19 0.19 0.25	6 7 8	96 48	0.75 0.75
7 8 9 10 11	0.75 1.5 2	0.19 0.25	7 8	48	0.75
8 9 10 11	1.5	0.25	8		
9 10 11	2			48	1
10 11		0.5	0		
11			9	128	0.75
	0.75	0.125	10	256	1
	1.5	0.25	11	96	0.75
12	1	0.25	12	256	1
13	1.5	0.25	13	48	0.75
14	2	0.38	14	64	1.5
15	2	0.38	15	32	0.25
16	1.5	0.38	16	32	0.75
17	2	0.25	17	24	0.75
18	1.5	0.25	18	24	1
MIC <sub>50</sub>	1.5	0.25	MIC <sub>50</sub>	56	0.75

In vitro activity of linezolid and tedizolid against methicillin-resistant

Table 2	Cumulative frequency of MIC <i>in vitro</i> of tedizolid and linezolid against isolates of methicillin-resistant <i>Staphylococcus aureus</i> (MRSA) and <i>cfr</i> -mediated methicillin- and linezolid-resistant <i>Staphylococcus aureus</i> (MLRSA).											
		MIC (mg/L)										
		0.125	0.25	0.5	1	2	4	8	>8			
MRSA	Linezolid				7	11						
	Tedizolid	2	11	5								
MLRSA	Linezolid							3	15			
	Tedizolid		2	2	13	1						

using the E-test (bioMérieux for linezolid and Liofilchem for tedizolid). Therefore, we sowed a bacterial lawn (0.5 McFarland) on Mueller-Hinton agar dishes and applied E-test strips. The dishes were incubated at 37°C for 18 hours.

#### RESULTS AND DISCUSSION

All MRSA isolates were susceptible to both oxazolidinones. However, when the strains were analyzed, the MIC $_{50}$  and MIC $_{90}$  of linezolid were 1.5 and 2 mg/L, respectively, whereas those of tedizolid were 0.25 and 0.4 mg/L. Analysis of the MLRSA strains (MIC $_{90}$  to linezolid  $\geq$  8 mg/L) revealed that the MIC $_{50}$  and MIC $_{90}$  of tedizolid remained at 0.75 and 1 mg/L. Despite the fact that the susceptibility cut-offs defined by the CLSI $^7$  and EUCAST $^8$  are 0.5 for tedizolid and 4 for linezolid, the concentrations obtained with tedizolid proved to be at least 2 dilutions lower, thus demonstrating greater *in vitro* activity (tables 1 and 2).

We used the E-test, a gradient diffusion method that is rapid, reliable, and widely applied in the routine of the clinical microbiology laboratory. The adequate correlation of this technique with broth microdilution (CLSI reference method) for testing susceptibility to *Staphylococcus* species has been demonstrated elsewhere<sup>9,10</sup>.

Tedizolid is a novel antibiotic from the oxazolidinone family, which until recently only comprised 1 member, linezolid. It is administered in the form of a phosphate ester, an inactive prodrug that is transformed into its active form via the action of plasma phosphates. Tedizolid is a stable drug within a wide range of pH values that binds to plasma proteins (70-90%) and has an oral bioavailability of more than 90%. Its high-affinity binding to plasma proteins and tendency toward high intracellular concentrations enable it to be administered in a single daily dose of 200 mg<sup>11</sup>. The advantages of tedizolid over linezolid are its more pronounced postantibiotic effect, improved posology, lower frequency of adverse effects, and a lower ability to select resistance mutations<sup>12</sup>.

Tedizolid was recently approved by the FDA and EMA for the treatment of infections of the skin and soft tissue, in which its liposolubility enabled it to reach high concentrations<sup>11</sup>. Linezolid was recently considered to be more widely distributed in tissue, even in areas that are difficult to access, such as necrotic tissue, bone, the blood-brain barrier, pulmonary epithelium, and alveolar macrophages<sup>13</sup>. However, the higher tissue concentrations reached by tedizolid, which are associated with lower myelotoxicity and drug interactions than linezolid<sup>14,15</sup>, make it an interesting alternative in the treatment of infections by MRSA and MLRSA.

Taken together, the *in vitro* activity we found and the characteristics of the drug in terms of diffusion and safety enable us to conclude that tedizolid could be an alternative in the complex and prolonged regimens used to treat infection by MRSA, even in patients whose resistance to linezolid is mediated by *cfr*.

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## **CONFLICTS OF INTEREST**

DLM is the Director of the Antibiotics Section of the Medical Department of MSD. FJC is a medical advisor on antimicrobial drugs for MSD, Pfizer, Astellas, Novartis, Gilead, and AstraZeneca.

#### REFERENCES

- López-Pueyo MJ, Barcenilla-Gaite F, Amaya-Villar R, Garnacho-Montero J. Multirresistencia antibiótica en unidades de críticos. Med Intensiva. 2011; 35: 41-53.
- Locke JB, Rahawi S, LaMarre J, Mankin AS, Shaw KJ. Genetic environment and stability of cfr in methicillin-resistant Staphylococcus aureus CM05. Antimicrob Agents Chemother 2012; 56: 332–40.
- LaMarre JM, Locke JB, Shaw KJ, Mankin AS. Low fitness cost of the multidrug resistance gene cfr. Antimicrob Agents Chemother 2011; 55: 3714–9.
- A. Shaw KJ, Poppe S, Schaadt R, Brown-Driver V, Finn J, Pillar CM et

- al. *In vitro* activity of TR-700, the antibacterial moiety of the prodrug TR-701, against linezolid-resistant strains. Antimicrob Agents Chemother 2008; 52: 4442–7.
- Locke JB, Finn J, Hilgers M, Morales G, Rahawi S, G C K, Picazo JJ et al. Structure-activity relationships of diverse oxazolidinones for linezolid-resistant *Staphylococcus aureus* strains possessing the cfr methyltransferase gene or ribosomal mutations. Antimicrob Agents Chemother 2010; 54: 5337–43.
- Sánchez-García M, De la Torre MA, Morales G, Peláez B, Tolón MJ, Domingo S et al. Clinical outbreak of linezolid-resistant Staphylococcus aureus in an intensive care unit. JAMA 2010; 303: 2260-4.
- CLSI. M100-S26: Performance Standards for Antimicrobial Susceptibility Testing. CLSI.org
- EUCAST Clinical Breakpoint Tables v. 6.0 (valid from 2016-01-01).
   Eucast.org
- Gómez-Garcés JL, López-Fabal F, Burillo A, Yolanda Gil A. Estudio comparativo de Wider, E-test y microdilución para la determinación de la sensibilidad a daptomicina y otros tres antimicrobianos de aislamientos clínicos de *Staphylococcus* spp. resistentes a meticilina y *Enterococcus* spp. Rev Esp Quimioter 2010; 23: 87-92.
- Nadarajah R, Post LR, Liu C, Miller SA, Sahm DF, Brooks GF. Detection of Vancomycin-Intermediate Staphylococcus aureus With the Updated Trek-Sensititre System and the MicroScan System. Comparison With Results From the Conventional Etest and CLSI Standardized MIC Methods. Am J Clin Pathol 2010; 133: 844-8.
- Moran GJ, Fang E, Corey GR, Das AF, De Anda C, Prokocimer P. Tedizolid for 6 days versus linezolid for 10 days for acute bacterial skin and skin-structure infections (ESTABLISH-2): a randomised, double-blind, phase III, non-inferiority trial. Lancet Infect Dis 2014;14:696-705. 10.1016/S1473-3099(14)70737-6.
- 12. Locke JB, Zurenko GE, Shaw KJ, Bartizal K. "Tedizolid for the Management of Human Infections: In Vitro Characteristics". Clin Infect Dis 2014; 58 (S1): S35–S42.
- Dryden MS. Linezolid pharmacokinetics and pharmacodynamics in clinical treatment. J Antimicrob Chemother 2011; 66 Suppl 4: iv7-iv15.
- Shorr AF, Lodise TP, Corey GR, De Anda C, Fang E, Das AF et al. Analysis of the phase 3 ESTABLISH trials of tedizolid versus linezolid in acute bacterial skin and skin structure infections. Antimicrob Agents Chemother 2015; 59: 864–71.
- 15. Flanagan S, McKee EE, Das D, Tulkens PM, Hosako H, Fiedler-Kelly J et al. Nonclinical and Pharmacokinetic Assessments To Evaluate the Potential of Tedizolid and Linezolid To Affect Mitochondrial Function. Antimicrob Agents Chemother 2015; 59: 178–85.